

# **Towards Reference Materials for Food Allergen and Gluten-Free Analysis**

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**MoniQA**  
[www.moniqa.org](http://www.moniqa.org)



SIXTH FRAMEWORK PROGRAMME



# MoniQA Association

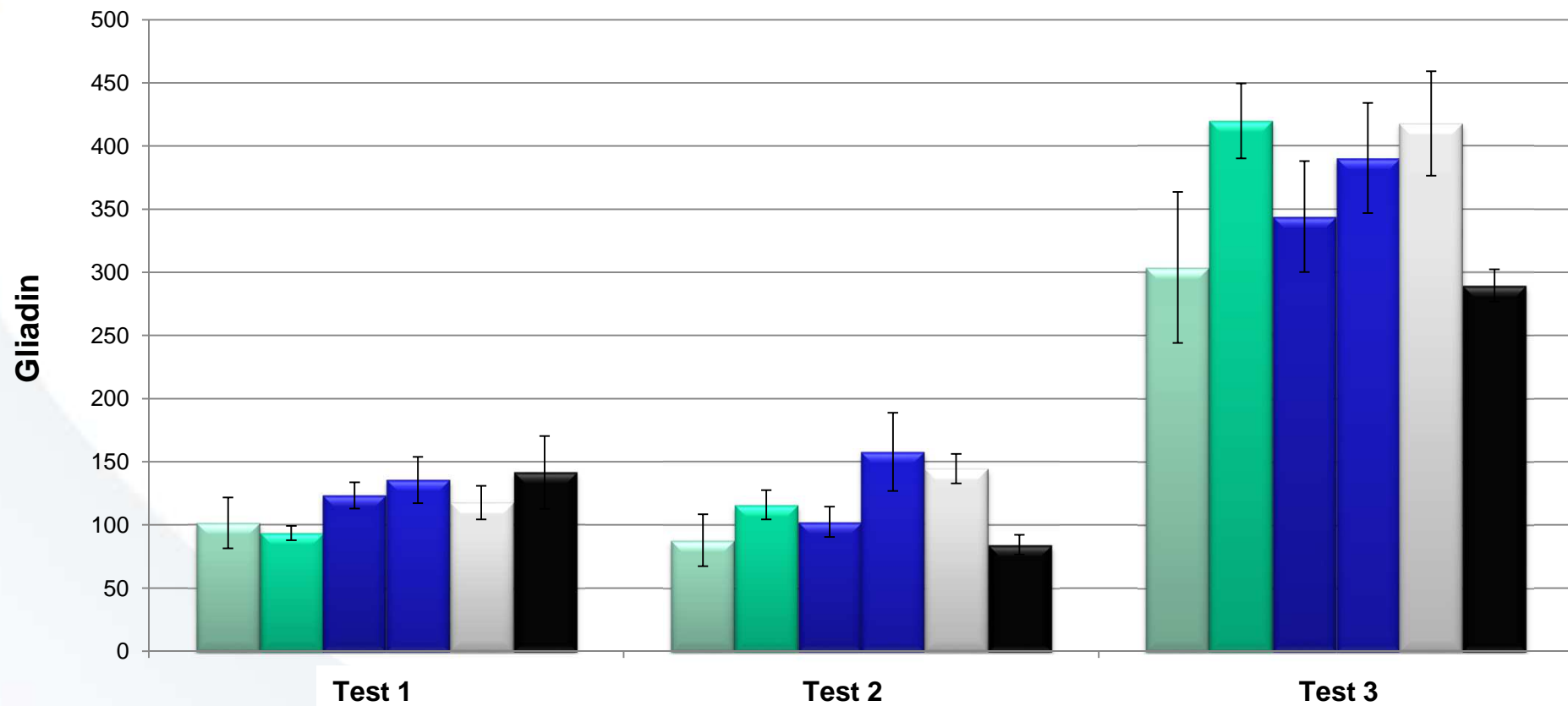
## MoniQA Global Food Safety Network

*...providing expertise and services for effective food safety management..*

*...for food manufacturers, food retailers, international trade, food analytical laboratories, food control and regulatory bodies*

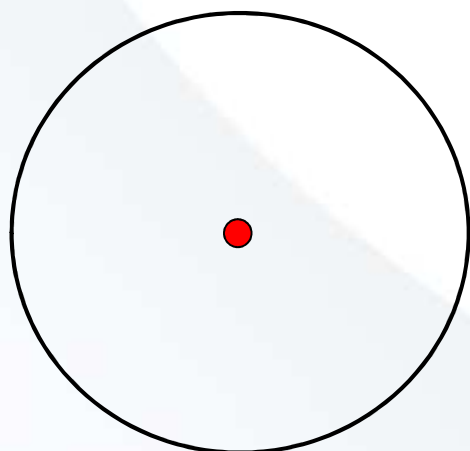
- Non profit-distributing association
- Established in July 2011
- Membership based
- Secretariat at ICC HQ in Vienna, Austria

# Example: Gliadin analysis by 3 different tests

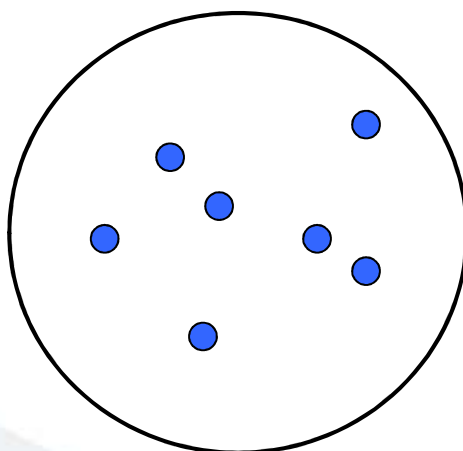


**How can I know  
that  
my analysis is correct?**

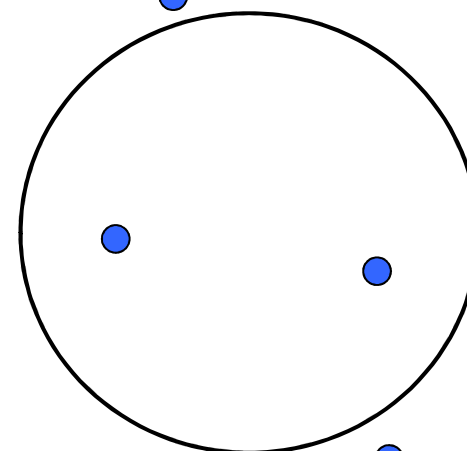
# Uncertainty – reliability of analytical results



Ideal

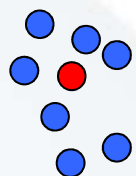


Reality

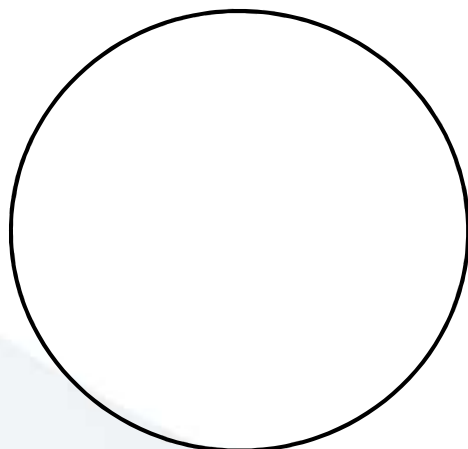


“Loser”

# Trueness – reliability of analytical results

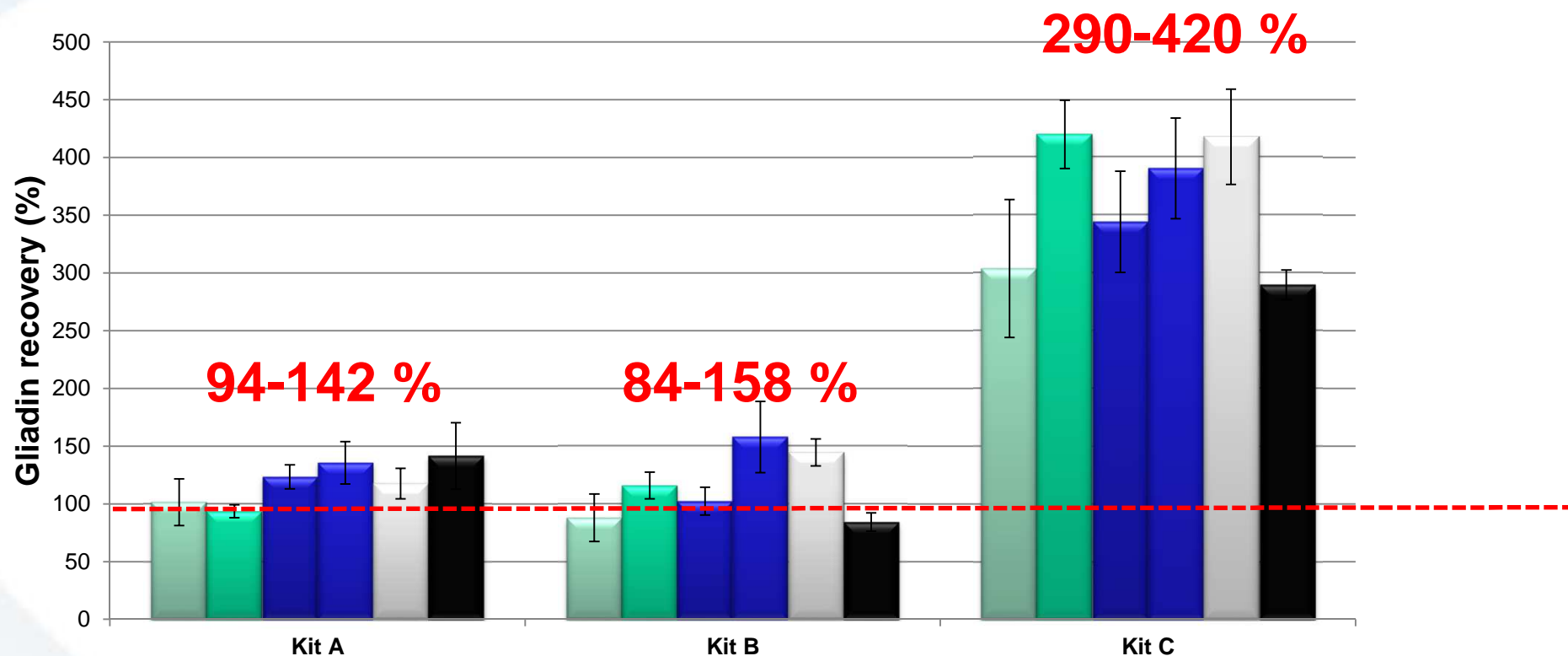


...looking good



...but way off

# Using a Reference Material



- Flour from one Hungarian wheat variety (MV-Magvas; Harvest year: 2011)
- Flour from one Hungarian wheat variety (MV-Magvas; Harvest year: 2012)
- Blended flour 1 (Mv-Magvas, MV-Mazurka, MV-Verbunkos, Dekan, Yumai-34; Harvest year: 2011)
- Blended flour 1 (Mv-Magvas, MV-Mazurka, MV-Verbunkos, Dekan, Yumai-34; Harvest year: 2012)
- Blended flour 2 (MV-Magvas, Bezostaja-1, Glenlea, Hereward, Soissons; Harvest year: 2012)
- PWG gliadin

*Hajas, L. , Tömösközi, S. , publication in preparation*



# MoniQA Working Group

# Food Allergen Reference Materials

**Coordinator:** Roland Poms (MoniQA, Austria)

**WG Committee:** Jupiter Yeung (**Nestle**, USA), Terry Koerner (**Health Canada**, Canada), Thomas Holzhauser (**Paul Ehrlich Institute**, Germany), Peter Koehler (**Prolamin WG**, Germany), Sigrid Haas-Lauterbach (**R-Biopharm**, Germany), Bert Popping (**AOAC**, USA), Joe Baumert (**FARRP**, USA), Robin Sherlock (**FACTA/Allergen Bureau**, Australia), Franz Ulberth (**European Commission, IRMM**, Belgium), Eric Garber (**FDA**, USA),...hosted by Kurt Johnson (**Trilogy Laboratories**, USA).

**Contributors and stakeholders:** Research institutes, test kit providers, method developers, food businesses, policy support, a.o.

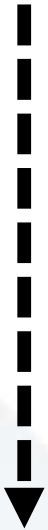


# Priority Allergen Testing Materials

- Basic materials (minimally processed), sufficiently characterised, as basis to produce incurred materials and extracts at different concentrations, model foods (e.g. rice cookie)
- Gluten/Gliadin, basic materials: flours of wheat, rye, barley, oat, rice
- Milk: dried skim milk powder
- Egg: dried full egg powder, dried egg white powder
- Hazelnut: flour
- Peanut: defatted peanut flour

# Characterization and Quality Control

## Allergenic food Commodities

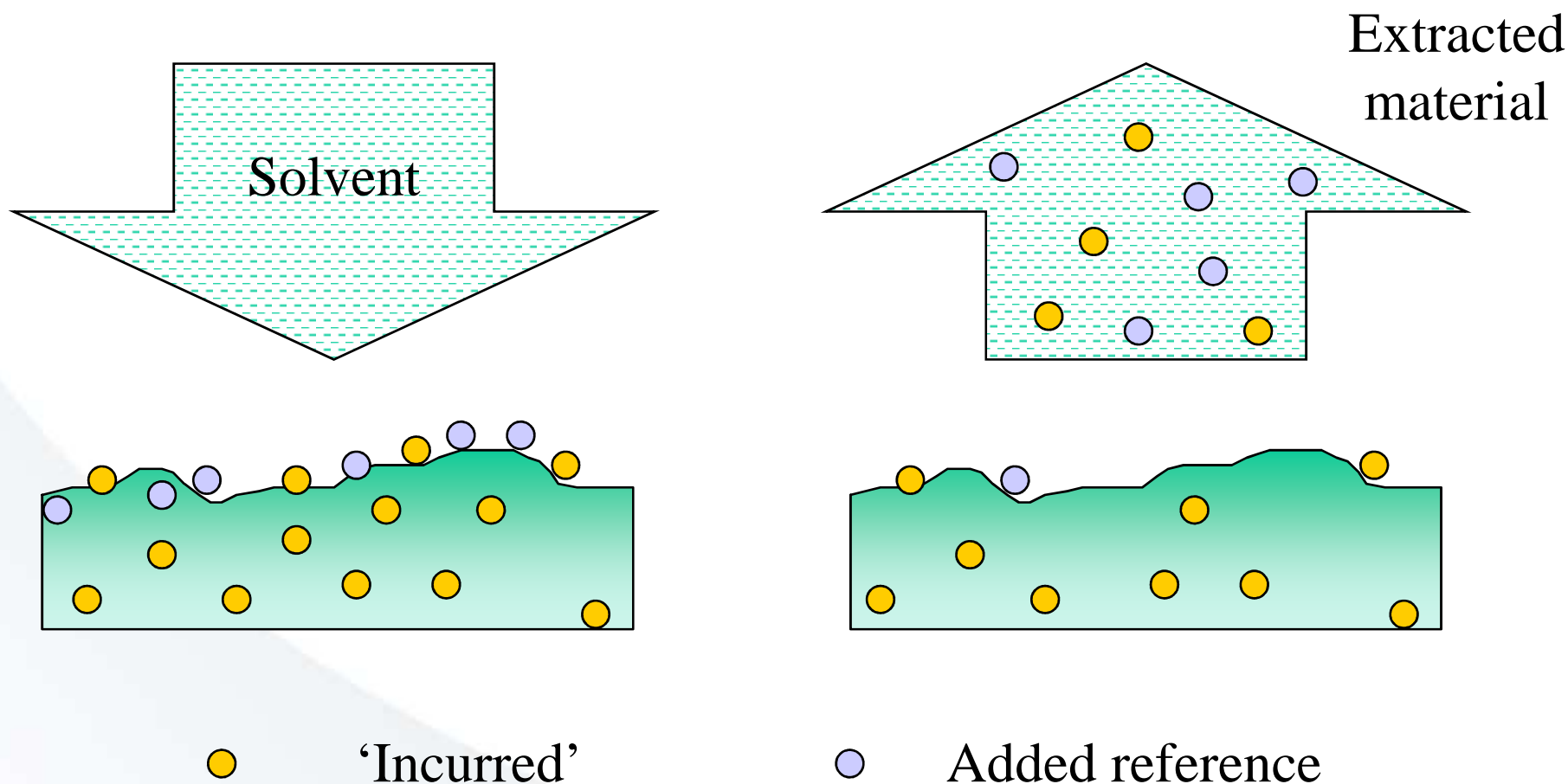


## Incurred RMs

Traceability documentation, provider's data  
Milling, sieving, packaging, particle size (40 µm)  
Protein, fat, moisture, ash analysis  
Protein profile by 2D electrophoresis  
Mass spec peptide profile (as available)  
Storage (4 ° C, -20 ° C, -80 ° C), stability testing

Production parameters documented  
Milling, sieving, packaging, particle size (40 µm)  
Protein, fat, moisture, ash contents  
Specific analysis by ELISA, PCR, LC-MS...  
Homogeneity (particle size, allergen analysis,  
Validated concentration (ring trial)  
Minimum sample intake  
Mass spec peptide profile (as available)  
Storage (4 ° C, -20 ° C, -80 ° C), stability testing  
Shelf life

# Incurred vs spiked controls



# MoniQA QC Material for Milk Allergen Analysis<sup>↑</sup>

- Skim Milk Powder was sourced September 2014 after the MoniQA Task Force Meeting at the last AOAC Annual Meeting held in Florida.
- Production of materials was outsourced to Trilogy Analytical Laboratory, Washington, Missouri, USA
- Stored at Trilogy at  $-16^{\circ}\text{C} \pm 1^{\circ}\text{C}$
- The material has analytical data for protein, fiber, moisture, carbohydrate and ash as well as a protein profile analysis.

# Certificate of Analysis

## issued by Trilogy Analytical Laboratory



**Reference Material**  
for the  
**Detection of Milk Proteins**  
Based on Skim Milk Powder  
**Product Code: SMP**  
**Art. No.: MQA 092014**

### Certificate of Analysis

| Analyte               | Lab 1                                | Lab 2                                | Lab 3                        | Average *       | 1 SD    | 2 SD range    |
|-----------------------|--------------------------------------|--------------------------------------|------------------------------|-----------------|---------|---------------|
|                       | Method Utilized for Determination    |                                      |                              |                 | (grams) | (grams)       |
| Ash                   | AOAC<br>923.03                       | AOAC<br>942.05                       | AOAC<br>930.30               | 7.56 g/100 g    | 0.15 g  | 7.26 – 7.86   |
| Carbohydrates         | Calculated                           | Calculated                           | Calculated                   | 51.61 g / 100 g | 0.52 g  | 49.97 – 52.65 |
| Fat                   | AOAC<br>989.05<br>Base<br>Hydrolysis | AOAC<br>954.02<br>Acid<br>Hydrolysis | AOAC<br>989.05<br>Mojonnier  | < 1.0 g / 100 g |         |               |
| Moisture              | AOAC<br>925.09                       | AOAC<br>925.09                       | SMEDP<br>15.111              | 5.01 g / 100 g  | 0.40 g  | 4.21 – 5.81   |
| Protein<br>(N x 6.38) | Kjeldahl<br>AOAC<br>979.09           | Kjeltec<br>AOAC<br>2001.11           | Kjeldahl<br>AOAC<br>991.20.1 | 35.39 g / 100 g | 0.20 g  | 34.99 – 35.79 |

\*this value represents the average of three independent external laboratory determinations. Each independent laboratory provided a single determination utilizing the methodology listed. The averaged result and 1 SD value are reported as well as the 2 SD range.

### Gel Electrophoresis Protein Identification

| Protein Identification | Band | Average | + SD |
|------------------------|------|---------|------|
| Unknown                | 1    | 2.2     | 0.14 |
| Unknown                | 2    | 0.60    | 0.09 |
| Unknown                | 3    | 1.5     | 0.13 |
| Unknown                | 4    | 0.76    | 0.12 |
| Lactoferrin            | 5    | 1.3     | 0.04 |
| Unknown                | 6    | 0.89    | 0.04 |
| Bovine Serum Albumin   | 7    | 1.1     | 0.03 |
| IgG Heavy Chain        | 8    | 2.5     | 0.05 |
| Unknown                | 9    | 1.9     | 0.05 |
| Alpha-casein           | 10   | 31.7    | 0.18 |
| Beta-casein, K-casein  | 11   | 32.0    | 0.39 |
| Para-K-casein          | 12   | 1.1     | 0.06 |
| Unknown                | 13   | 1.5     | 0.18 |
| Beta Lactoglobulin     | 14   | 14.3    | 0.09 |
| Unknown                | 15   | 0.56    | 0.05 |
| Alpha-lactalbumin      | 16   | 4.6     | 0.18 |
| Unknown                | 17   | 1.0     | 0.06 |

Method: SDS Slab Gel Electrophoresis by method of Laemmli, (UK Nature 227: 680-685, 1970) as described by Burgess-Cassler et. al. (Clin. Chem. 35:2297-2304, 1989; second dimension) using a 12% acrylamide slab gel (125 mm length x 150 mm width x 0.75 mm thickness) overlaid with a 25 mm stacking gel. Electrophoresis was performed using a 15 mAmp/gel for about 3.5 hours at which time bromophenol blue front had migrated to the end of the slab gel. The gel was stained with Coomassie blue, destained in 10% acetic acid until a clear background was obtained, then dried between cellophane sheets. The stained gel was digitized over the appropriate optical density range using a calibrated GE Healthcare Image Scanner III. Stain density in individual protein bands as a percentage of stain per line were quantified using Phoretix software.

**Certification Date:** September 2014

**Expiry date:** Sept 2019

Signed:

Carrie Maune for Trilogy Analytical Laboratory



# Samples available for further testing

- Packets contain an overfill of 5 grams and are contained in a zippered foil pouch
- Stored at -16° C
- Planned contingent to last for about 5 years with 7000 packets per commodity
- About 1000 packets ready
- About 20 laboratories have signed up for testing by ELISA and LC-MS, other?



# **MoniQA QC Material Blank for Spiking Experiments and Basis of Incurred**

- Extensive sourcing was done to secure rice flour, potato starch, xanthan gum, baking powder and baking soda with allergen levels below detection for Total Milk, Gluten, Soy, Egg, peanut and Hazelnut.
- Material was pre-sampled and tested in various laboratories to confirm allergen results.
- Large amounts of the tested “clean” lots were ordered and repackaged in large foil zippered packages.
- All products are stored at Trilogy under -16° C conditions.



# MoniQA QC Material

## Rice Cookie Incurred with Milk

- 1) Dry Skim Milk was weighed, dissolved in water and incorporated in cookie dough mixtures at starting concentrations of **100 ppm and 500 ppm**, respectively
- 2) **Baking at 165° C for 20 min**
- 3) **Grinding and sieving at mesh 35/40**
- 4) Production of lower concentrations by blending blank and incurred material to desired content: **0, 5, 10, 50 ppm SMP** corresponding to approximately **0, 1.8, 3.5, 16.8 mg SMP protein/kg incurred cookie**.
- 5) Packaging and storage
- 6) Testing for homogeneity and recovery (by ELISA) at various stages
- 7) Inter-laboratory testing by ELISA and MassSpec

# MoniQA QC Material

## Rice Cookie Incurred with Milk

### 100 ppm SMP

98.6 ppm SMP  
34.89 ppm milk  
protein

1/3 + 2/3 blank

2/3 + 1/3 blank

undiluted

11.63  
ppm  
milk  
protein  
**Low**  
**A1**

23.26  
ppm  
milk  
protein  
**Low**  
**A2**

34.89  
ppm  
milk  
protein  
**Low**  
**A3**

### 500 ppm SMP

498.3 ppm SMP  
176.35 ppm milk  
protein

1/3 + 2/3 blank

2/3 + 1/3 blank

undiluted

58.78  
ppm  
milk  
protein  
**High**  
**A1**

117.56  
ppm  
milk  
protein  
**High**  
**A2**

176.35  
ppm  
milk  
protein  
**High**  
**A3**

# Preliminary Batch Results – “Total Milk”

## BASIC HOMOGENEITY

|                | ELISA #1     | ELISA #2     |
|----------------|--------------|--------------|
| Low A1         | 7.6 %        | 1.6 %        |
| Low A2         | 12.7 %       | 12.8%        |
| Low A3         | 8.0 %        | 7.5%         |
| High A1        | 5.3 %        | 8.7%         |
| High A2        | Outlier      | 4.2%         |
| High A3        | <u>4.6 %</u> | <u>12.7%</u> |
| <b>Average</b> | <b>7.6 %</b> | <b>7.9 %</b> |
|                |              |              |

## BASIC RECOVERY

|                | ELISA #1      | ELISA #2      |
|----------------|---------------|---------------|
| Low A1         | 92.0 %        | 62.5 %        |
| Low A2         | 75.2 %        | 76.1 %        |
| Low A3         | 65.6 %        | 73.3 %        |
| High A1        | 83.4%         | 71.1 %        |
| High A2        | 55.4 %        | 70.5 %        |
| High A3        | <u>63.7 %</u> | <u>62.2 %</u> |
| <b>Average</b> | <b>72.6 %</b> | <b>69.3 %</b> |
|                |               |               |

# Gluten/Gliadin RM

- 1. Raw materials:** Wheat, rye, barley and oat, rice, corn
- 2. Format:** Commodity flour
- 3. Incurred materials:** different concentrations of wheat/rye/barley flour in rice cookie
- 4. Selection of variety:**
  - setting criteria
  - quantitative
  - qualitative

# Considerations

- Reference Material (RM): as representative as possible for the multitude of wheat cultivars grown around the world
- ? Importance of geographic origin and other variables: cultivar, climate, year of harvest
  - Selection of a representative cultivar
- ? White flour or whole wheat flour
  - White flour (better stability)
- ? One cultivar or a mixture of several cultivars
  - One cultivar (problems with a mixture are homogeneity and high complexity)
- ? Isolation of gluten fractions (similar to PWG-gliadin)
  - Possible in cooperation with the Prolamin Working Group

# RM Development for GLU/GLI analysis

## Collection of seeds

**35 wheat varieties**

(Australian, Austrian, Canadian, Chinese, German, Hungarian, etc.)

**2 Rye varieties**

**4 Barley varieties**

## Production of white flour

## Characterisation

### NIR Spectrum

Infratec Grain Analyzer 1241

(850-1048 nm)

- NIRSystems 6500 (400-1098, 1100-2498 nm)

**Wet and dry gluten**  
(Glutomatic System)

**1D SDS PAGE**

**RP- and SE-HPLC**

**Western Blotting**

## Production of flour

## Incurred material

**ELISA**

**2D SDS PAGE**

**Criteria for selection of variety?**

Median Gliu/Glu rate?

Median Protein/Gluten content?

Mostly used variety

# Selection criteria

Based on means of all measurements (35 wheat flours/mixtures)  $\pm$  standard deviation

## Quantitative Criteria

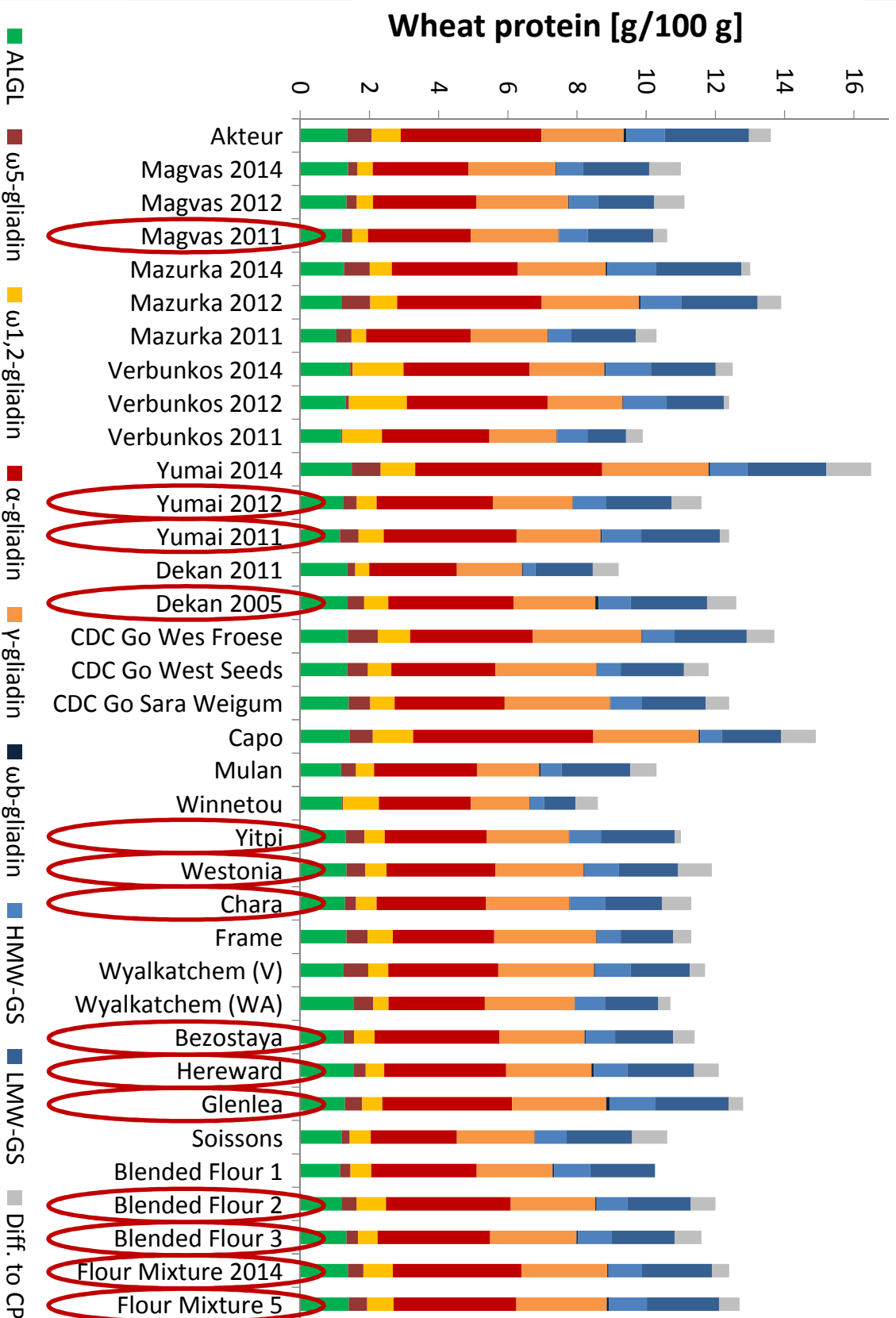
|   |                   |                 |
|---|-------------------|-----------------|
| Crude protein content (Dumas method):       | $11.8 \pm 1.5 \%$ | (10.3 - 13.3 %) |
| Ratio prolamin/glutelin:                    | $2.6 \pm 0.5$     | (2.1 - 3.1)     |
| Ratio LMW/HMW:                              | $2.1 \pm 0.6$     | (1.5 - 2.7)     |
| Content of $\omega$ 5-gliadin:              | $3.9 \pm 1.7 \%$  | (2.2 - 5.6 %)   |
| Content of $\omega$ 1,2-gliadin:            | $6.5 \pm 2.4 \%$  | (4.1 - 8.9 %)   |
| Ratio $\alpha$ -gliadin/ $\gamma$ -gliadin: | $1.4 \pm 0.2$     | (1.2 - 1.6)     |

## Qualitative Criteria

|                    |  |
|--------------------|--|
| HMW-GS:            | at least 5 HMW-GS                                    |
| $\gamma$ -gliadin: | no pronounced peaks after an elution time > 19.5 min |

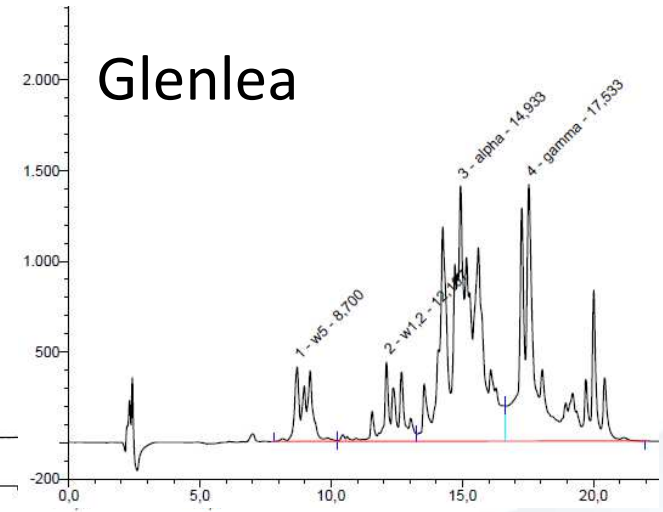
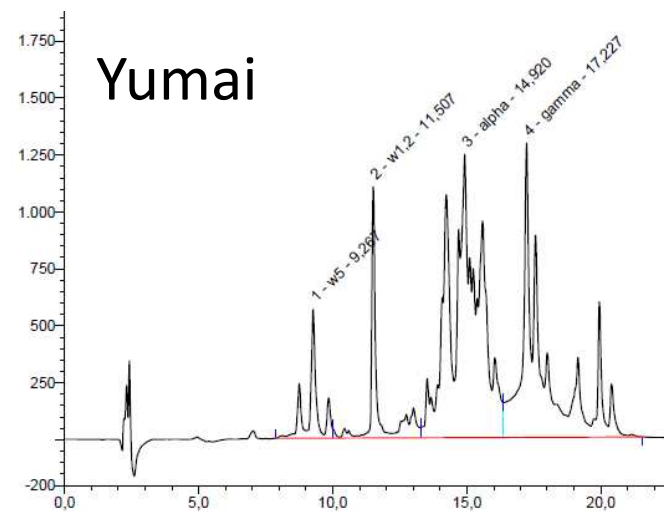
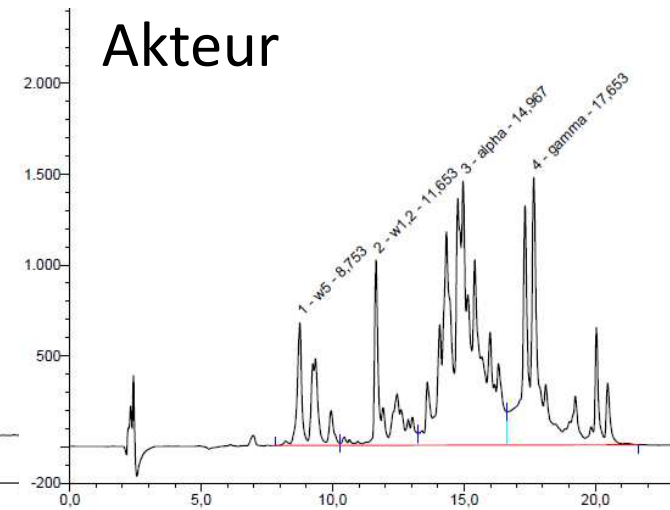
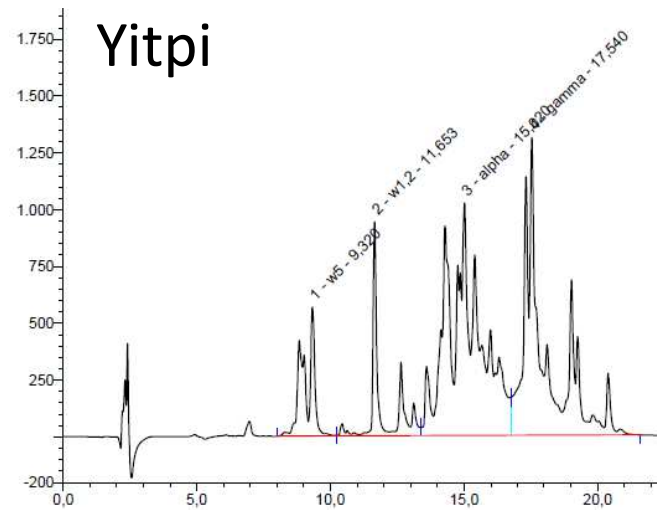
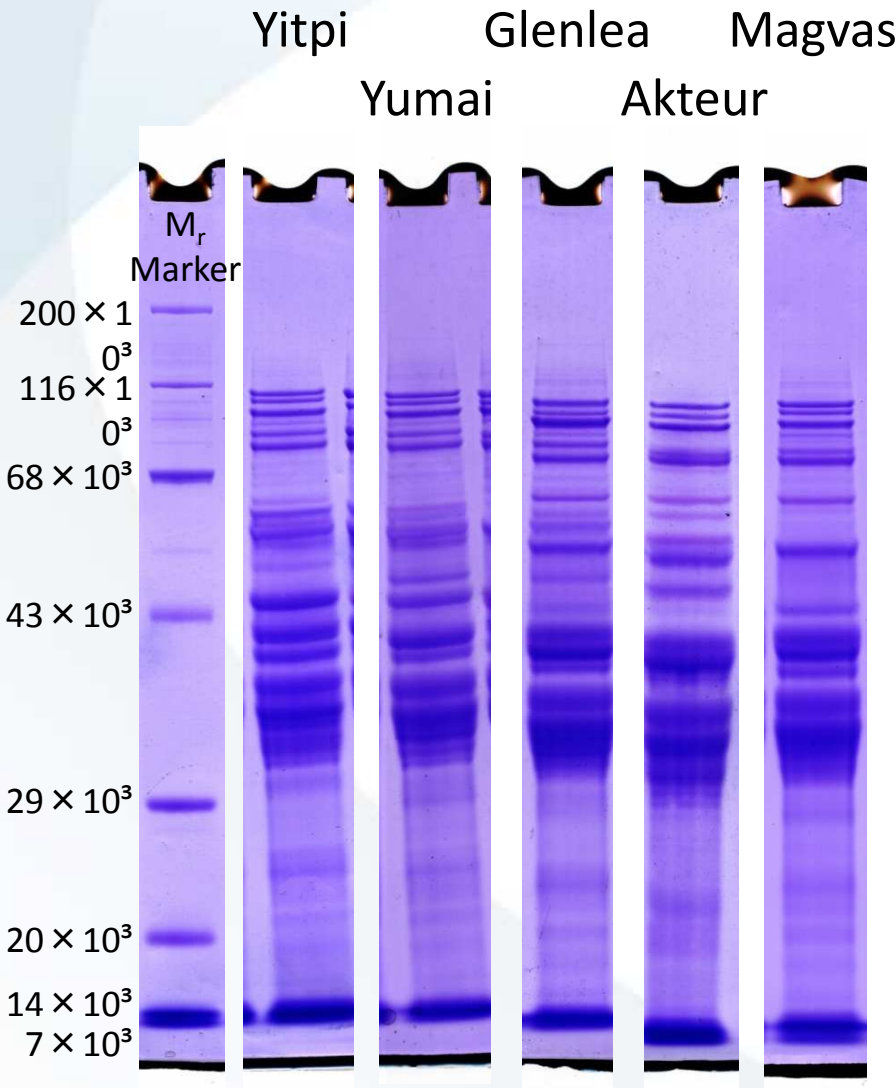


# RP-HPLC of wheat flours



Quantitative criteria fulfilled

# 1D SDS-PAGE + RP-HPLC (Gliadins)



# Summary

- **Reference Materials** are needed for food allergen and Gluten-free analysis
- An **international consortium** led by MoniQA took the initiative
- **Dried skim milk** is available, characterized for food allergen analysis
- **Blank** matrix material based on a gluten free rice cookie is available
- **Different concentrations** of milk in a rice cookie (incurred) to become available by the end of 2015: 0, 5, 10, 50 ppm SMP
- **Inter-laboratory testing** of milk and blank in October/November 2015
- **Wheat and gluten free** materials being characterized, first availability of materials in 2016



# FOOD AND HEALTH – RISKS AND BENEFITS

## INNOVATIVE TECHNOLOGIES FOR FOOD QUALITY AND SAFETY MANAGEMENT



**Next Task Force Meeting: prior to PWG in Tulln, Austria  
8 October 2015, 16:00-19:00**

5<sup>TH</sup> MONIQA INTERNATIONAL CONFERENCE 2015  
16-18 SEPTEMBER 2015, PORTO, PORTUGAL

**MoniQA**  
Association

 **iceta**



# Get involved?!

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